





"Tet and Cre" dependent genetic logic circuit and its application in mammalian cells





Outline

- Introduction
- Project & Modeling
- Results & Summary
- Human practice & safety
- Acknowledgements

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Introduction



All cells! Internal or from environment? Produce appropriate cellular response

What we hope is to construct genetic logic circuits like this.



BMC Syst Biol. 2013 Jul 24;7:67. doi: 10.1186/1752-0509-7-67.

Introduction

Previous work in both prokaryotic and eukaryotic cells



Moon et al. Nature. 2012 Nov 8;491(7423):249-53.



Garber et al. Nat Chem Biol. 2014 Mar;10(3):203-8.

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Project -- Brainstorming



Project -- Motif searching





We choose Bi-Fan motif as the blueprint of our circuit design.

If output signals of A/B/C/D regions can be separated, theoretically we can construct any common logic gate!



Project -- Basic Elements



Project -- Overview









• knock down --- shRNA undergo microRNA genesis and produce siRNA, knock down target gene



• sequence shorten?! --- TM activate Cre recombinase, excise sequence between loxP



Project -- Design Conclution





Project -- Modeling



Project -- Application

Application: cell fate decision



Use this logic circuit ABCD gene replaced with certain genes to induce cell transdifferentiation and reprogramming.

Brief introduction for application

• A OSKM (four transcription factors linked with T2A, used to somatic cell reprogramming to iPSc)



Takahashi K, Yamanaka S.. cell, 2006, 126(4): 663-676.

• B Sox2 (used to somatic cell reprogramming to NSc)



Ring K L, Tong L M, Cell stem cell, 2012, 11(1): 100-109.

• C shRNA for PTB (PTB gene knock down for somatic cell transd if ferentiation to neuron)



Xue Y, Ouyang K, Huang J, et al Cell, 2013, 152(1): 82-96.



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Results & Summary

• 1. Testing our parts

Useful information is on

Fudan 2014 iGEM Team Parts

http://Parts.igen.org

Favorite Fudan 2014 iGEM Team Parts								
< < Name	Type	Description	Designer	Length	\$			

Fudan 2014 iGEM Team Parts Sandbox Edit								Edit	
\$	\$	Name 🔶	Туре	Description	\$	Designer	\$	Length	\$
1	W	BBa_K1440000	Regulatory	Part2_pTRE promoter with cFP and ribozyme		Xuanye Cao		1	1291
	W	BBa_K1440066	Regulatory	reverse CMV->reverse RFP->pTRE->shRNA		Xuanye Cao			1192
	W	BBa_K1440088	Regulatory	long parts consist of part2 and part3		Xuanye Cao			2491

Use These three parts to test our three working systems: Cre-ERTII system, Tet-on system Ribozyme system

All the bio-bricks we used to test our systems are all included in the parts we submitted and registered.

Validation of Cre-TM system

• Result of testing Cre-TM system





[TM]

[TM]

/ml

10umol

<u>5umol/</u> ml

<u>2umol/</u> ml

blood cell counting chamber

Efficiency Calculation: GFP-positive cell numbers/ Total cell numbers





cotransfection with Cre-ERTII and part2 EGFP plasmid with loxp sites/ +TM 2umol 36hr



cotransfection with Cre-ERTII and part2 EGFP plasmid with loxp sites/ +TM_5umol_36hr



cotransfection with Cre-ERTII and part2 EGFP plasmid with loxp sites/ +TM_10umol_36hr

[TM] Oumol/ <u>ml</u>

Control

GFP



cotransfection with Cre-ERTII and part2 EGFP plasmid with loxp sites/ no TM 36hr



cotransfection with Cre-ERTII and part2 EGFP plasmid with loxp sites/ +TM_1umol_36hr



Validation of Cre-TM system



amoxi

Our Cre-TM system Works!

Validation of tet-on shRNA expression system

Result of testing tet-on shRNA system



figure 1₽

Validation of tet-on shRNA expression system

Result of testing tet-on shRNA system





Conclusion: Our tet-on shRNA system Works!

Validation of Ribozyme system

• Result of testing ribozyme system



CMV mCherry

Conclusion: Our Ribozyme Works!

Use tet-on system to get iPSc!

C(Dox)=1ug/ml





T2A

pTRE

P2A

A. The single clone of iPS cells

B. Using the Alkaline Phosphatase kits(Sigma) to detect the iPS cells



C&D. The tet-induced iPS cells with the OSKM plasmid.

Use tet-on shRNA system to get Neuron

- (b) using tet-on shRNA expression system to test neuron differentiation
- c(dox)=1ug*t*m l

- Conclusion:
- We successfully knock down PTB protein, with 3 designed shRNA.





qPCR for PTB knock down result

Results & Summary

• Summary:

(a) We successfully construct three parts for testing three systems which are most important to our design. And we get the quantitative data of three systems. However, because the limitation of time and many mistakes we made before, we have not assembled three parts together yet.

(b) We use three parts to test our application: cell fates' decision. We successfully get iPSc cell clone. For neuron and iNSc cell culture, although we have the help from Tongji University, we still cannot get the right result with our working-successfully parts because of limitation of time.

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Human practice & Outreach

- We help several teams, and also get their help;
- We join in the meets up in Taiwan
- We hold events like Bacteria Painting Competition to spread the spirit of iGEM and synthetic biology.
- We construct our own student association for iGEM competition.
- http://



 $\boldsymbol{\cdot}$ We offer help to Tongji University

Http://





With Tongji University

They help us with the protocol of cell transdifferentiation and cell reprogramming.



With Zhejiang University

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 We introduce our project to them, and give some advice to their project



With NYU Shanghai

 We help them build their "next-year" team and wet lab for iGEM



Meets up in NCTU-Formosa



We give a presentation about our project.





We talk about our poster to the other 20 schools

Certification!

Construct our own student's association for iGEM

Bertalanffy club





Bacteria Painting Competition



Safety

	Advantages	Disadvantag es
iPSc	strong self- renew ability	oncogenicity
	strong differentiat ion potential	low efficiency
somatic cell conversion	no oncogenicity	no self- renew abilities
	high efficiency	poor differentiat ion potential

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Thanks to them for their teaching 0 0~